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Complete Genome Sequence of Bacillus cereus Bacteriophage BCP78

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Bacillus cereus is generally found in soil habitats, and it contaminates a wide variety of foods, causing food poisoning with symptoms such as vomiting and diarrhea. To develop a novel biocontrol agent to inhibit this pathogen, bacteriophage BCP78 belonging to the *Siphoviridae* family was isolated from a fermented food sample. Here we announce the complete genome sequence of BCP78, which may be useful for understanding its inhibition mechanism against *B. cereus*, and describe major findings from the genome annotation.

B*acillus cereus* is a soil bacterium and is generally propagated to vegetables and foods (4, 6, 7). It sometimes causes severe food poisoning with symptoms such as nausea, vomiting, and diarrhea (4, 13). Because bacteriophage treatment was allowed for use in foods by the U.S. FDA (2), it would be useful to ensure food safety from *B. cereus* contamination (9, 14). However, genomic characterization for its inhibition mechanism has not been conducted yet. The *B. cereus* bacteriophage BCP78 was isolated from a fermented food, and it belongs to the *Siphoviridae* family, efficiently inhibiting *B. cereus* and several strains of *B. subtilis* in various vegetables and foods (data not shown).

Alkaline lysis method (15) was used to isolate the phage genomic DNA, and it was sequenced using the Genome Sequencer FLX (GS-FLX) Titanium by Macrogen, Seoul, South Korea. The assembly of quality filtered reads was performed using 454 Newbler 2.3 assembler, and the prediction of open reading frames (ORFs) and their confirmation were conducted using the Glimmer 3.02 (3), GeneMark.hmm (10), and FgenesV softwares (Softberry, Inc., Mount Kisco, NY), respectively. Conserved protein domain analysis of predicted ORFs was also carried out using the BLASTP (1) and InterProScan programs (16). Comparative codon preference analyses of the *B. cereus* and phage BCP78 genomes were performed using GCUA program (11).

The complete circular genome of B. cereus phage BCP78 showed a 156,176-bp length with a GC content of 39.86%, 227 ORFs, and 18 tRNAs, suggesting the first phage genome targeting B. cereus in the Siphoviridae family with the largest number of tRNAs. It is intriguing that this genome encodes many extra tRNAs, suggesting that they probably help translation of the host mRNAs or they may be involved in translation of the phage mRNAs (8). Comparative codon usage analysis of B. cereus strains (AH187 and ATCC 14597) and the phage showed different preferences in phenylalanine, asparagine, and serine, explaining a possible role for extra tRNAs in the phage genome. This phage genome encodes structural and packaging proteins, such as a major capsid protein, a prohead protease, minor structural proteins, baseplate proteins, a portal protein, a terminase, a tail sheath protein, tail lysins, and tail fiber proteins. This genome encodes two copies of tail lysins, probably involved in the infection mechanism (5), and an autolysin and a holin, probably involved in cell lysis (12). In addition, this genome encodes many DNA manipulation proteins (DNA polymerases, helicases, a primase and a recombinase) and nuclease (DNA nucleases and endo- and exonucleases). Furthermore, there are three putative sigma factors in this genome, suggesting that they are probably related to preferential transcription of the phage genes rather than the host genes. The complete-genome analysis of *B. cereus* phage provides novel information about *B. cereus*-targeting phage.

Nucleotide sequence accession number. The complete genome sequence of *B. cereus* phage BCP78 is available in GenBank under accession number JN797797.

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REFERENCES

- 1. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J. Mol. Biol. 215:403–410.
- 2. Cairns BJ, Payne RJH. 2008. Bacteriophage therapy and the mutant selection window. Antimicrob. Agents Chemother. 52:4344-4350.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679.
- Drobniewski FA. 1993. Bacillus cereus and related species. Clin. Microbiol. Rev. 6:324–338.
- Kenny JG, McGrath S, Fitzgerald GF, van Sinderen D. 2004. Bacteriophage Tuc2009 encodes a tail-associated cell wall-degrading activity. J. Bacteriol. 186:3480–3491.
- 6. Kim SK, et al. 2009. Prevalence and toxigenic profiles of *Bacillus cereus* isolated from dried red peppers, rice, and sunsik in Korea. J. Food Prot. 72:578–582.
- King NJ, Whyte R, Hudson JA. 2007. Presence and significance of *Bacillus cereus* in dehydrated potato products. J. Food Prot. 70:514–520.
- Kunisawa T. 1992. Synonymous codon preferences in bacteriophage T4: a distinctive use of transfer RNAs from T4 and from its host *Escherichia coli*. J. Theor. Biol. 159:287–298.
- Lee WJ, Billington C, Hudson JA, Heinemann JA. 2011. Isolation and characterization of phages infecting *Bacillus cereus*. Lett. Appl. Microbiol. 52:456–464.

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- Lukashin AV, Borodovsky M. 1998. GeneMark.hmm: new solutions for gene finding. Nucleic Acids Res. 26:1107–1115.
- McInerney JO. 1998. GCUA: general codon usage analysis. Bioinformatics 14:372–373.
- O'Flaherty S, Ross RP, Coffey A. 2009. Bacteriophage and their lysins for elimination of infectious bacteria. FEMS Microbiol. Rev. 33: 801–819.
- 13. Schoeni JL, Wong AC. 2005. *Bacillus cereus* food poisoning and its toxins. J. Food Prot. **68**:636–648.
- Shin H, Bandara N, Shin E, Ryu S, Kim K-P. 2011. Prevalence of *Bacillus cereus* bacteriophages in fermented foods and characterization of phage JBP901. Res. Microbiol. 162:791–797.
- Wilcox SA, Toder R, Foster JW. 1996. Rapid isolation of recombinant lambda phage DNA for use in fluorescence *in situ* hybridization. Chromosome Res. 4:397–398.
- 16. Zdobnov EM, Apweiler R. 2001. InterProScan-an integration platform for the signature-recognition methods in InterPro. Bioinformatics 17: 847–848.