Journal of Virology

Complete Genome Sequence of Cronobacter sakazakii Bacteriophage CR3

Hakdong Shin, Ju-Hoon Lee, Yeran Kim and Sangryeol Ryu *J. Virol.* 2012, 86(11):6367. DOI: 10.1128/JVI.00636-12.

	Updated information and services can be found at: http://jvi.asm.org/content/86/11/6367
	These include:
REFERENCES	This article cites 12 articles, 4 of which can be accessed free at: http://jvi.asm.org/content/86/11/6367#ref-list-1
CONTENT ALERTS	Receive: RSS Feeds, eTOCs, free email alerts (when new articles cite this article), more»

Information about commercial reprint orders: http://jvi.asm.org/site/misc/reprints.xhtml
To subscribe to to another ASM Journal go to: http://journals.asm.org/site/subscriptions/



Complete Genome Sequence of *Cronobacter sakazakii* Bacteriophage CR3

Hakdong Shin,^a Ju-Hoon Lee,^b Yeran Kim,^a and Sangryeol Ryu^a

Department of Food and Animal Biotechnology, Department of Agricultural Biotechnology and Center for Agricultural Biomaterials) Seoul National University, Seoul, South Korea, and Department of Food Science and Biotechnology, Kyung Hee University, Yongin, South Korea

Due to the high risk of *Cronobacter sakazakii* infection in infants fed powdered milk formula and the emergence of antibiotic-resistant strains, an alternative biocontrol agent using bacteriophage is needed to control this pathogen. To further the development of such an agent, the *C. sakazakii*-targeting bacteriophage CR3 was isolated and its genome was completely sequenced. Here, we announce the genomic analysis results of the largest *C. sakazakii* phage known to date and report the major findings from the genome annotation.

Cronobacter sakazakii is an opportunistic food-borne pathogen that often contaminates powdered infant milk formula, vegetables, and fruits and causes septicemia, meningitis, and necrotizing enterocolitis in neonates (3, 6, 9). It has recently attracted public attention due to the extremely high risk to infants fed contaminated formula, with 50 to 80% fatality rates (5, 9). The high resistance of *C. sakazakii* to unusually dry conditions supports a high survival rate in powdered infant formula (4). However, the emergence of antibiotic-resistant *C. sakazakii* strains has limited the use of antibiotics to control this pathogen (12), suggesting that the development of alternative biocontrol agents, like bacteriophage, is urgently needed. Here, a novel *C. sakazakii* bacteriophage, CR3, was isolated from an environmental sample and its genome was completely sequenced.

Genomic DNA was isolated using a standard alkaline lysis method (10) and sequenced using GS-FLX Titanium by Macrogen, Seoul, South Korea. The quality-filtered reads were assembled using Newbler 2.3, and the prediction of open reading frames (ORFs) was performed using GeneMarkS (1), Glimmer3 (2), and FgenesV (Softberry, Inc., Mount Kisco, NY). Transfer RNAs were predicted using tRNAscan-SE (8), and conserved protein motif analyses of the predicted ORFs were conducted using Inter-ProScan (11). Comparative codon preference analyses of the *C. sakazakii* BAA-894 (7) and phage CR3 genomes were carried out using CodonW 1.4.4 in the MOBYLE portal website (Pasteur Institute, Paris, France).

The complete genome of C. sakazakii phage CR3, belonging to the Myoviridae family, is 149,273 bp in length with a GC content of 50.95%, 265 ORFs, and 18 tRNAs, indicating the largest genome among C. sakazakii bacteriophages to date. Interestingly, this phage genome contains many tRNAs, and comparative codon preference analyses between the phage and C. sakazakii BAA-894 showed different codon preferences for valine, serine, alanine, lysine, asparagine, arginine, and glycine, suggesting that these extra phage tRNAs may play a role in the translation of phage mRNA, not host mRNA (data not shown). The genome of phage CR3 encodes structure/packaging proteins (major capsid protein, head stabilization/decoration protein, tail fiber proteins, tail fiber assembly protein, tape measure protein, and terminase), DNA manipulation proteins (DNA polymerases, DNA methylases, DNA primase, DNA helicase, DNA ligase, DNA methyltransferase, and endonucleases), and many additional functional proteins, such as a thymidylate synthase and a cell wall hydrolase, SleB. Although this genome encodes many ORFs, most of them are hypothetical proteins (84.5%), probably due to insufficient information about *C. sakazakii* phage genes in the GenBank database. Phage CR3 has two copies of tail fiber proteins targeting flagella of *C. sakazakii*, experimentally confirmed by the fact that CR3 did not infect the flagellum deletion mutant (data not shown). While this phage genome does not encode endolysin for host lysis, it encodes a cell wall hydrolase, SleB, suggesting that this protein may be involved in the host lysis. The complete genome analysis of *C. sakazakii* phage CR3 provides further information about *C. sakazakii* phages and extends the potential for application of phage CR3 as a natural biocontrol agent to control *C. sakazakii*.

Nucleotide sequence accession number. The complete genome sequence of *C. sakazakii* bacteriophage CR3 is available in GenBank under accession number JQ691612.

ACKNOWLEDGMENT

This work was supported by a National Research Foundation of Korea (NRF) grant funded by the Ministry of Education, Science and Technology (no. 20090078983).

REFERENCES

- 1. Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res. 29:2607–2618.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679.
- 3. Drudy D, Mullane NR, Quinn T, Wall PG, Fanning S. 2006. *Enterobacter sakazakii*: an emerging pathogen in powdered infant formula. Clin. Infect. Dis. 42:996–1002.
- Gurtler JB, Kornacki JL, Beuchat LR. 2005. Enterobacter sakazakii: a coliform of increased concern to infant health. Int. J. Food Microbiol. 104:1–34.

Received 14 March 2012 Accepted 15 March 2012
Address correspondence to Sangryeol Ryu, sangryu@snu.ac.kr.
H.S. and J.-H.L. contributed equally to this article.
Copyright © 2012, American Society for Microbiology. All Rights Reserved.
doi:10.1128/JVI.00636-12

- 5. Healy B, et al. 2010. *Cronobacter* (*Enterobacter sakazakii*): an opportunistic foodborne pathogen. Foodborne Pathog. Dis. 7:339–350.
- Kandhai MC, Reij MW, Gorris LGM, Guillaume-Gentil O, van Schothorst M. 2004. Occurrence of Enterobacter sakazakii in food production environments and households. Lancet 363:39–40.
- Kucerova E, et al. 2010. Genome sequence of Cronobacter sakazakii BAA-894 and comparative genomic hybridization analysis with other Cronobacter species. PLoS One 5:e9556.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964.
- 9. Nazarowec-White M, Farber JM. 1997. Enterobacter sakazakii: a review. Int. J. Food Microbiol. 34:103–113.
- Wilcox SA, Toder R, Foster JW. 1996. Rapid isolation of recombinant lambda phage DNA for use in fluorescence in situ hybridization. Chromosome Res. 4:397–398.
- Zdobnov EM, Apweiler R. 2001. InterProScan—an integration platform for the signature-recognition methods in InterPro. Bioinformatics 17: 847–848.
- 12. Zhou XF, Gao JX, Huang YJ, Fu SZ, Chen HY. 2011. Antibiotic resistance pattern of *Klebsiella pneumoniae* and *Enterobacter sakazakii* isolates from powdered infant formula. Afr. J. Microbiol. Res. 5:3073–3077.

6368 jvi.asm.org Journal of Virology